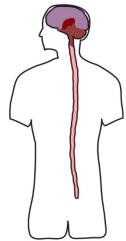
Optical control of neural ablation in zebrafish as a model for secondary injury mechanisms

Karen Mruk, Patrick A. Piza, and James K. Chen Departments of Chemical and Systems Biology, Developmental Biology, and Chemical Engineering

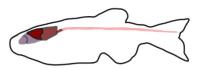
Stanford, University, Stanford, CA

Behavioral consequences of neuronal loss



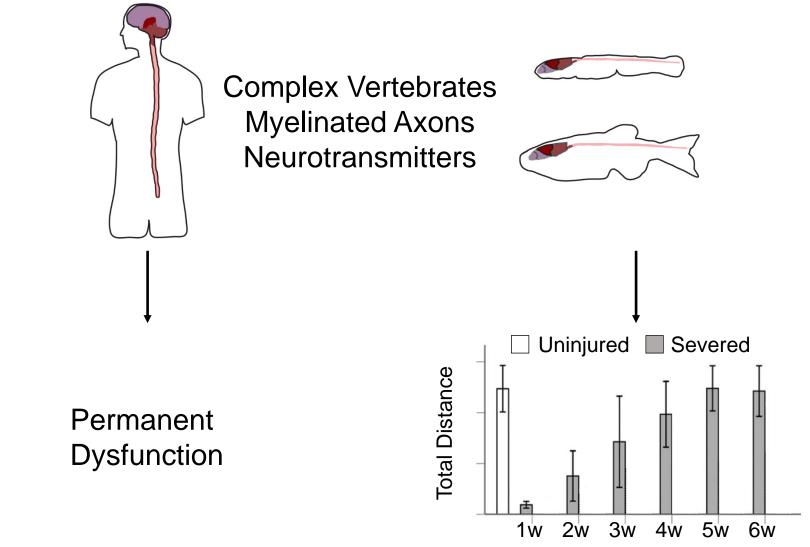
Complex Vertebrates Myelinated Axons Neurotransmitters







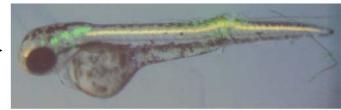
Behavioral consequences of neuronal loss



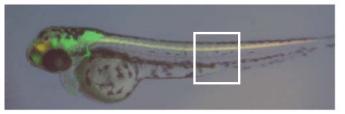
Adapted from: Yu, Y and Schachner, M. Eur J Neurosci. 2013



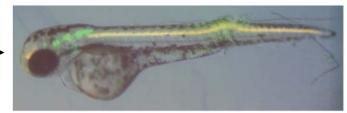
Manual Stabbing

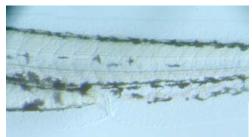


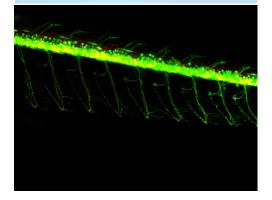


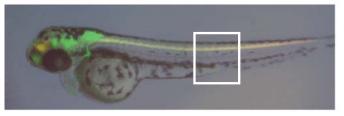


Manual , Stabbing

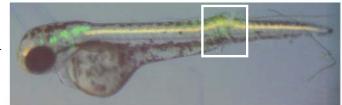


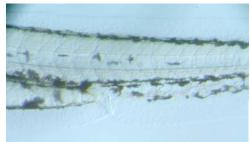


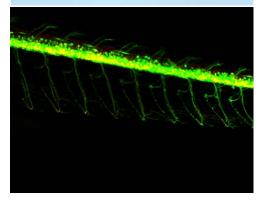


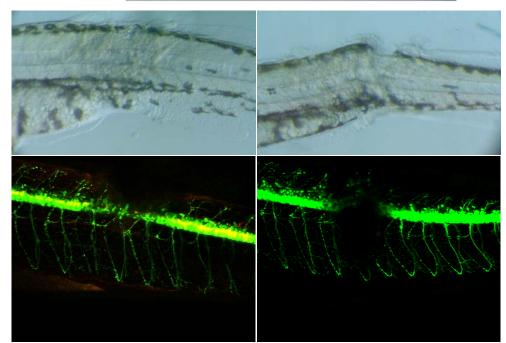




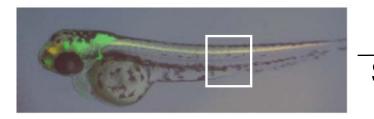


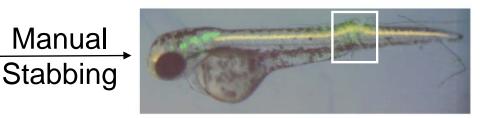




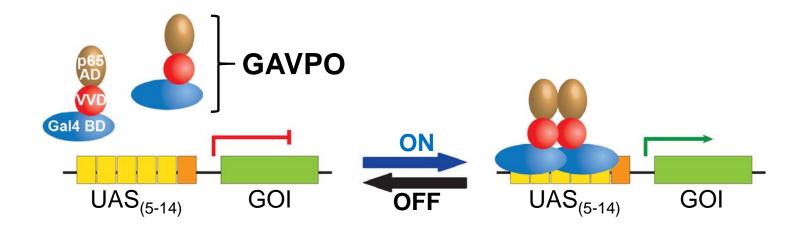


Less reproducible Extraneous damage Higher mortality

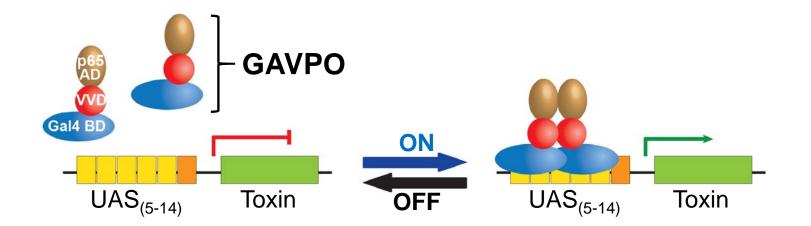




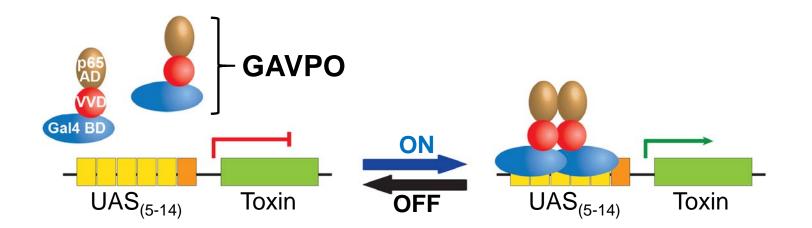
Can we use an optogenetic approach to reproducibly kill neurons as a model for spinal cord injury and degeneration?



Wang et al. Nature Methods 2012

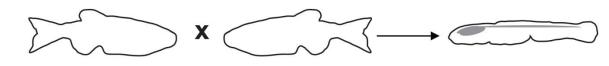






- Does GAVPO drive transcription in zebrafish?
- Can we model neural death with toxins?
- □ Can we induce neural death using the GAVPO system?

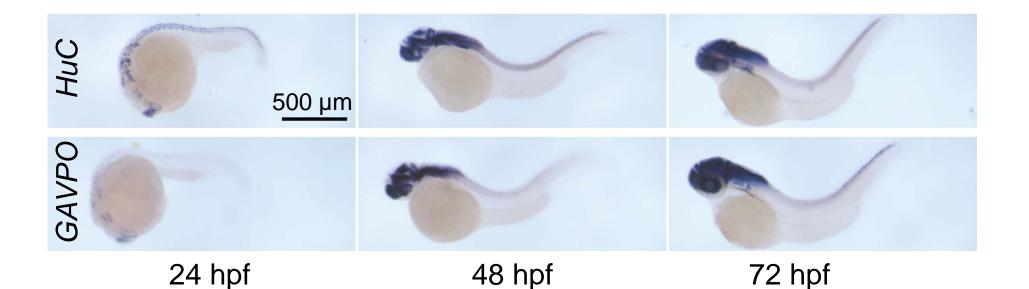
Generating and characterizing transgenic GAVPO lines



WT

Tg(*HuC:GAVPO*)

Fix and stain for mRNA expression



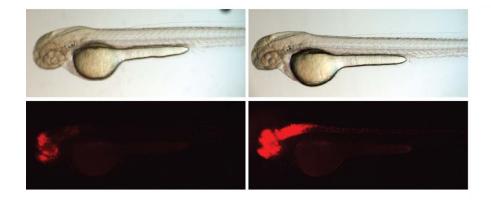
n = 20-25 larvae for each condition

GAVPO-induced expression increases with irradiation duration and intensity



Tg(HuC:GAVPO) Tg(UAS:mCherry)

Irradiate at 60 hpf Image at 72 hpf



20 min 120 min Global Irradiation 3 mW/cm²

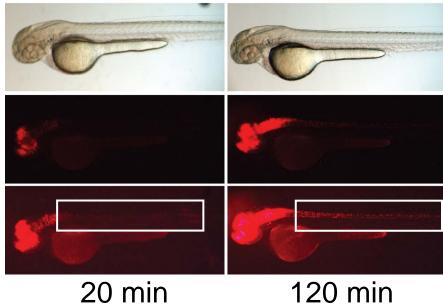
n = 25-30 larvae for each condition

GAVPO-induced expression increases with irradiation duration and intensity



Tg(HuC:GAVPO) Tg(UAS:mCherry)

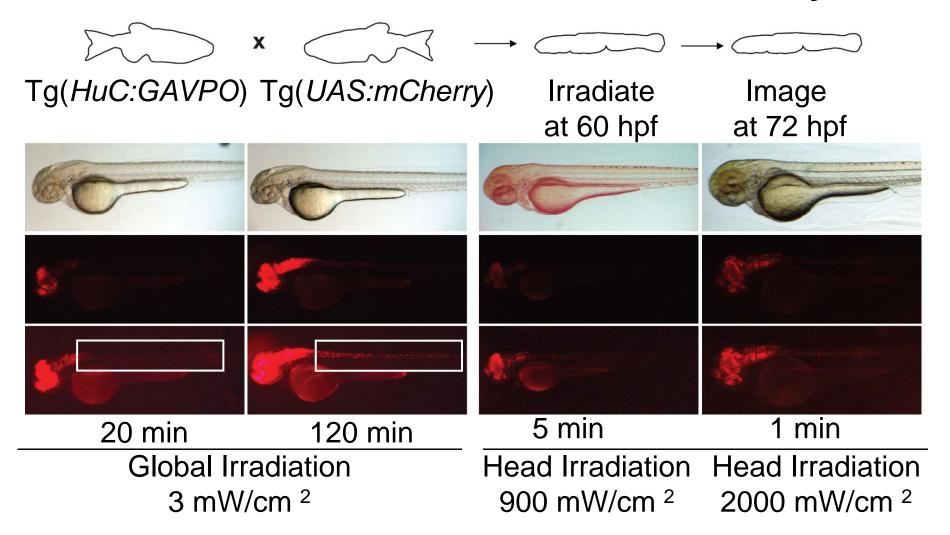
Irradiate at 60 hpf Image at 72 hpf



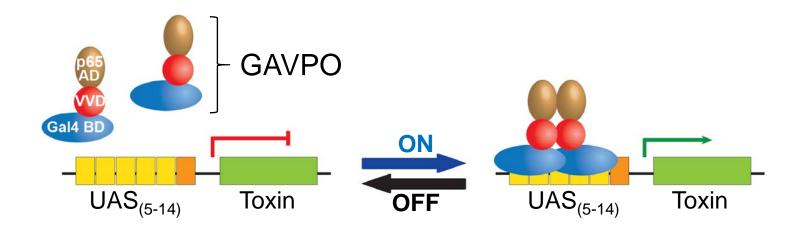
Global Irradiation 3 mW/cm²

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GAVPO-induced expression increases with irradiation duration and intensity

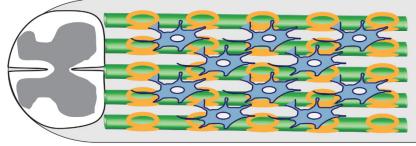


n = 25-30 larvae for each condition

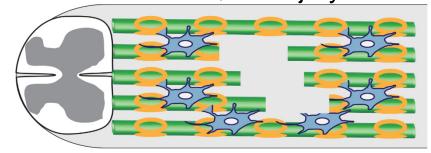


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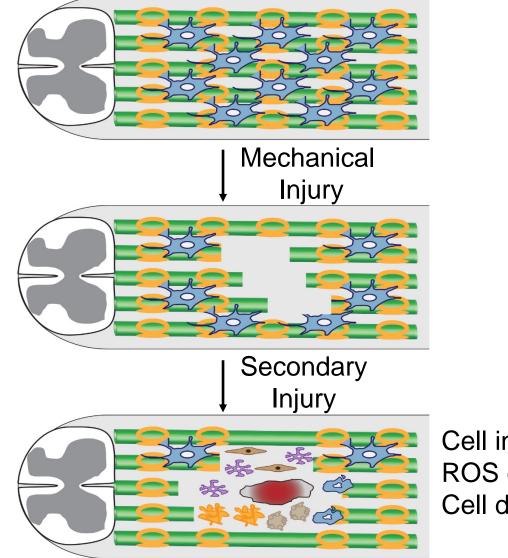
Modeling secondary injury via toxin-mediated ablation



↓ Mechanical ↓ Injury

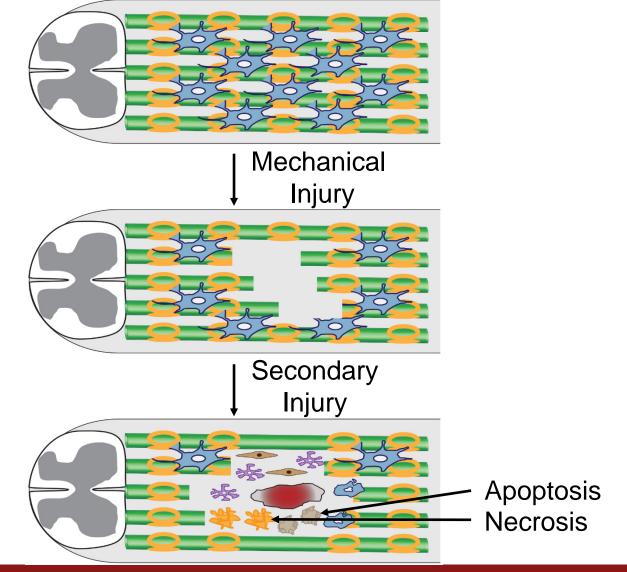


Modeling secondary injury via toxin-mediated ablation

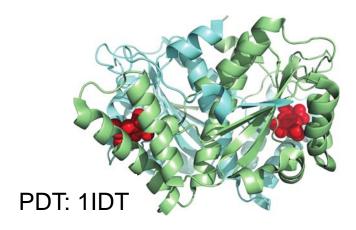


Cell infiltration ROS generation Cell death

Modeling secondary injury via toxin-mediated ablation

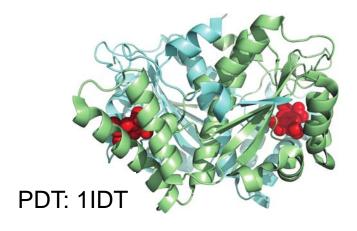


Modeling neuronal loss via development of toxin-mediated ablation

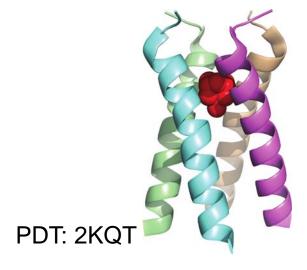


- Nitroreductase
 - Enzyme converts a prodrug (metronidazole) into a cytotoxic compound
 - Gold standard for zebrafish
 - Disadvantage slow acting

Modeling neuronal loss via development of toxin-mediated ablation



- Nitroreductase
 - Enzyme converts a prodrug (metronidazole) into a cytotoxic compound
 - Gold standard for zebrafish
 - Disadvantage slow acting



- M2
 - Ion channel from flu
 - Can be blocked with FDA approved drug (rimantadine)
 - Never used in zebrafish
 - Advantage fast acting

Generating and characterizing Tg(UAS:M2) lines









48 hpf



No drug control

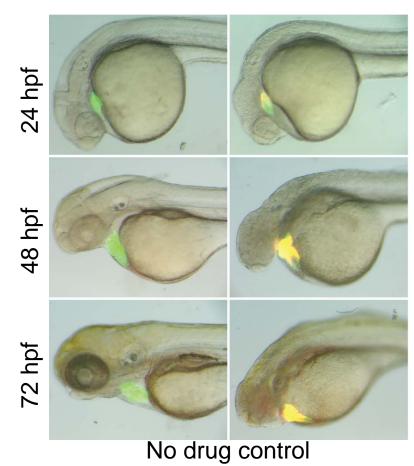
n = 30-50 larvae for each condition

Generating and characterizing Tg(UAS:M2) lines





Tg(*α-tub:Gal4VP16; UAS:M2*)



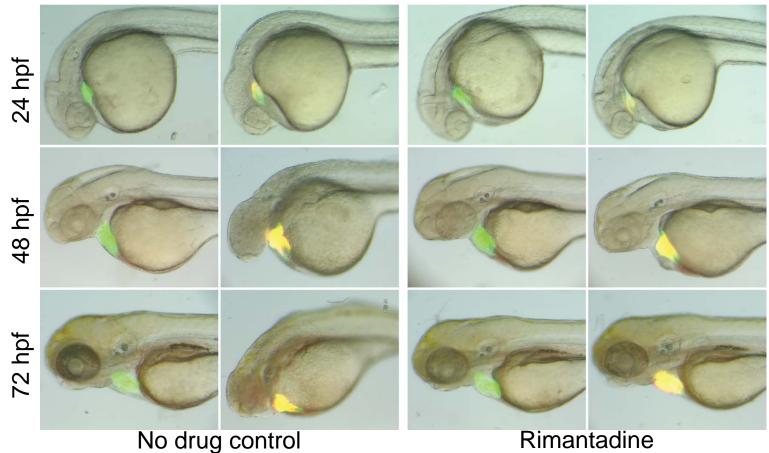
n = 30-50 larvae for each condition

Generating and characterizing Tg(UAS:M2) lines





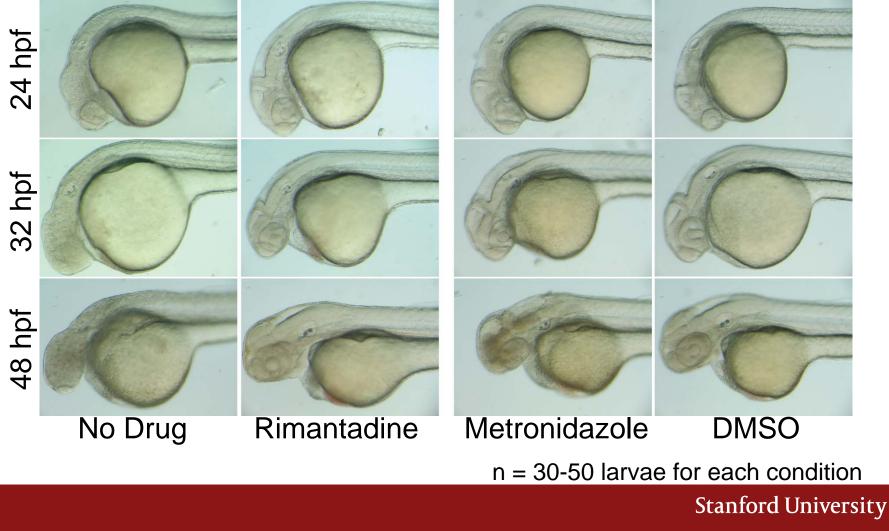


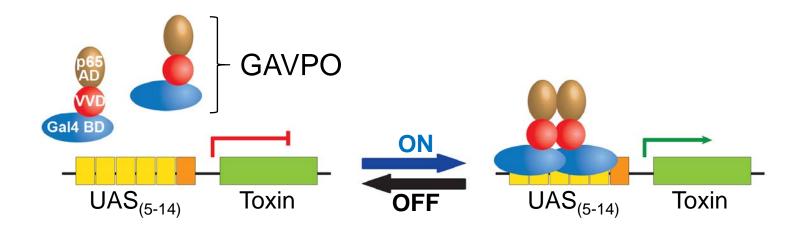


Rimantadine n = 30-50 larvae for each condition

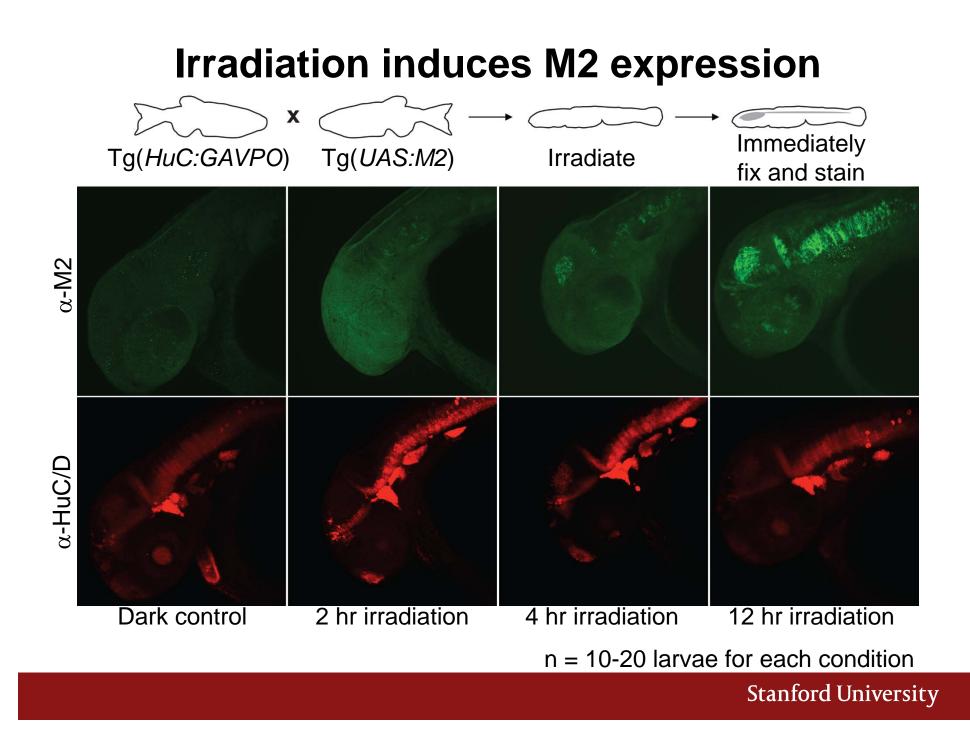
M2-induced defects are more pronounced than NTR-mediated ablation

Tg(α -tub:Gal4VP16; UAS:M2) Tg(α -tub:Gal4VP16; UAS:NTR)

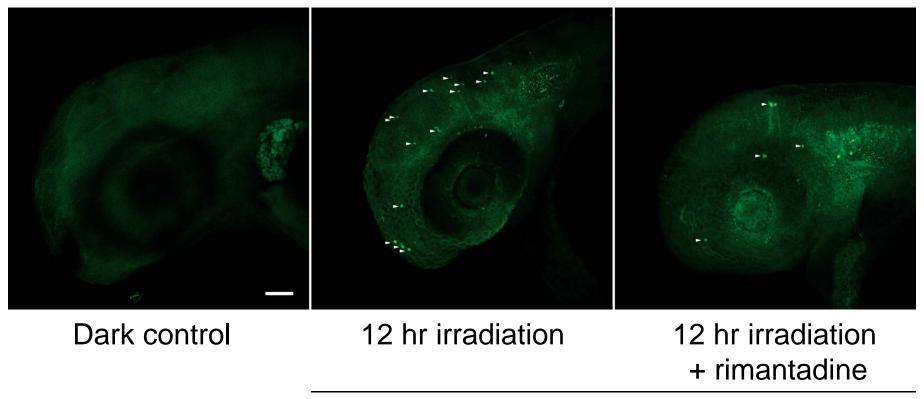




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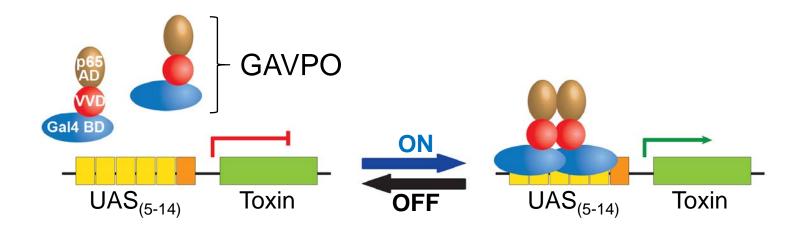


Irradiation induces activation of Caspase-3 which can be attenuated with rimantadine



Global irradiation (470-nm light 3 mW/cm²)

n = 10-20 larvae for each condition

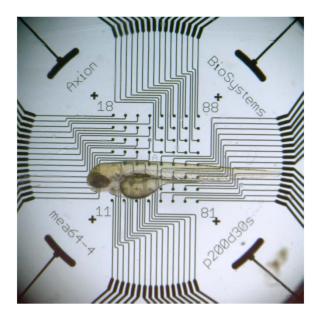


- ✓ Does GAVPO drive transcription in zebrafish?
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How does the nervous system response to optogenetic ablation compare to manual injury?

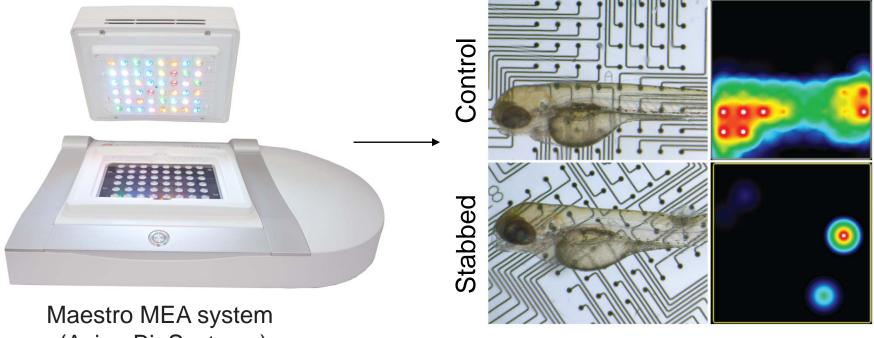
Comparing neural ablation to traditional spinal cord injury in zebrafish





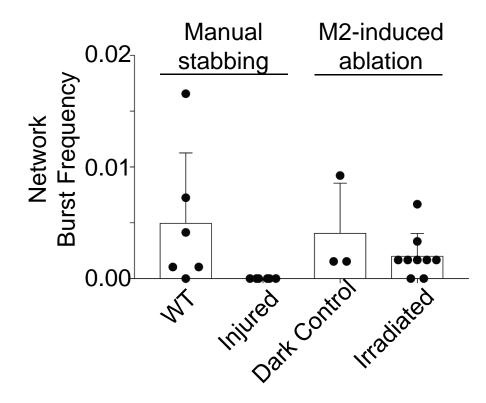
Maestro MEA system (Axion BioSystems)

Comparing neural ablation to traditional spinal cord injury in zebrafish

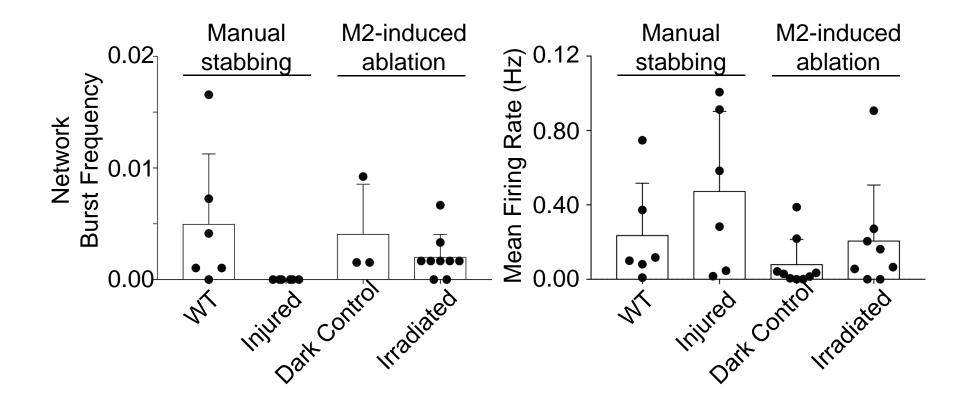


(Axion BioSystems)

Injury reduces network bursting and increases firing frequency



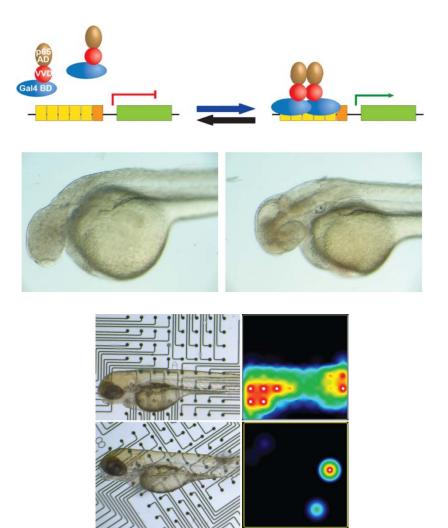
Injury reduces network bursting and increases firing frequency



n = 3-10 larvae for each condition

Summary and future directions

- Developed transgenic zebrafish for light-inducible gene expression
- Used multiple toxins to model secondary injury
- Studying neuronal circuitconnectivity and physiology and SCI pathology



Acknowledgements

- Chen Laboratory
 - James Chen
 - Moe Alnaqid
 - Patrick Piza
- Plant Laboratory
 - Giles Plant
- Axion Biosystems
 - Stacie Chvatal
 - Emily Matheu



