OPTIMIZATION OF A HUMAN STEM CELL DERIVED NEURON/ASTROCYTE CO-CULTURE SYSTEM FOR SEIZURE LIABILITY ASSESSMENT USING MICROELECTRODE ARRAYS



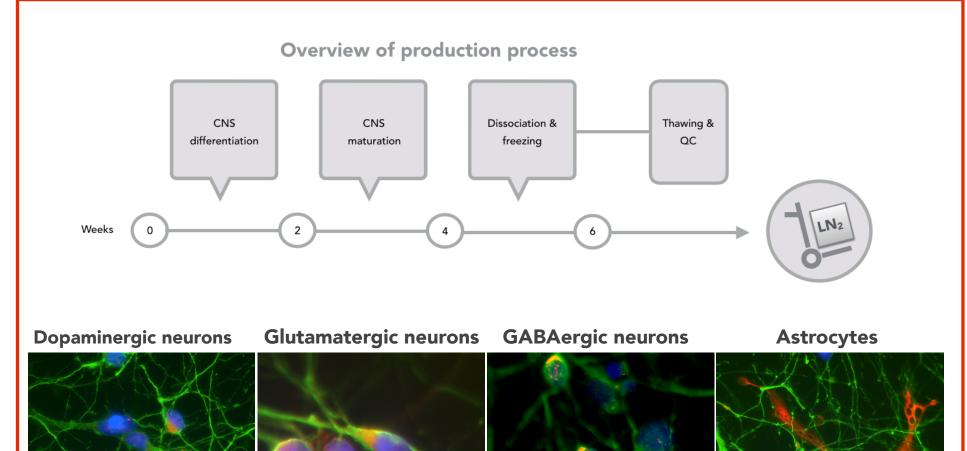
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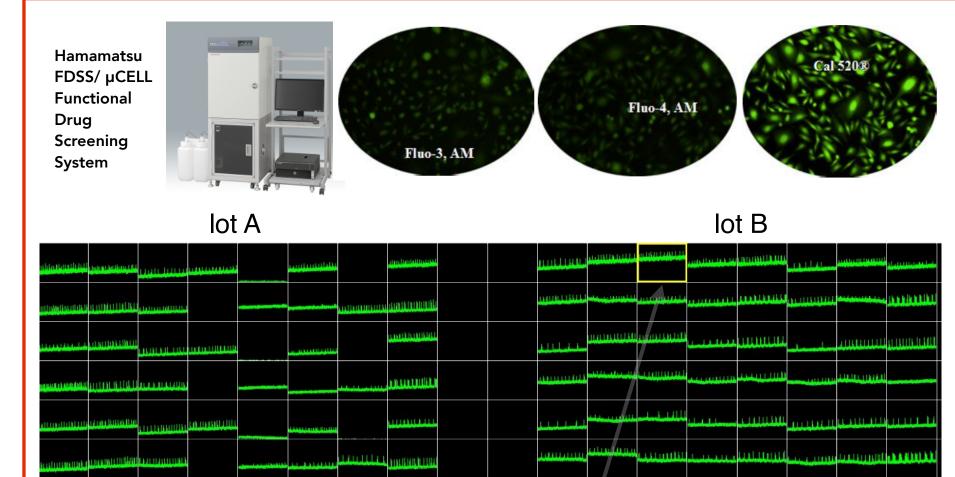
BACKGROUND

Historically, animal EEG studies have been the standard for preclinical assessment of drug induced seizures. Furthermore, in a typical ex vivo study, cortical neurons derived from rat forebrain must be extracted and cultured on microelectrode arrays (MEA) for roughly 4 weeks before mature functional network activity can be utilized for seizure assessment. With recent advances in human stem cell technologies, iPS-derived neurons can provide spontaneous electrical activity closely resembling that of murine ex vivo preparations. Here, using Axion Maestro MEAs, the electrophysiological function was compared amongst three different iPS-derived neuronal subtypes in the presence and absence of astrocytes. As with ex vivo preparations, we found that astrocytes are indeed necessary to provide iPS neurons with the physiological co-culture environment required for mature network level activity. Once network level activity was achieved (typically 2-3 weeks), co-cultures were exposed to 12 different compounds having a variety of seizure-related, anti-seizure, or neurotoxic activity (e.g. GABA A, K+ channel, Na+ channel, muscarinic ACh, glycine, D2 receptor, & MAO block). Though "time to assay readiness" was different, sensitivity to these compounds were similar for the three neuronal populations. Importantly, these co-culture models all demonstrated good predictivity within the 12 drug set and allowed for significantly faster "assay ready" culture times than typical murine ex vivo preparations. In conclusion, human iPS neurons + astrocytes provide a number of advantages over current models for seizure liability and anti-epileptic drug screening efforts and should be further explored to develop a more comprehensive library to better understand their predictivity for drug induced seizures.

Ncardia hiPSC-derived neurons show increased translatability with unlimited supply potential ideal for HTS



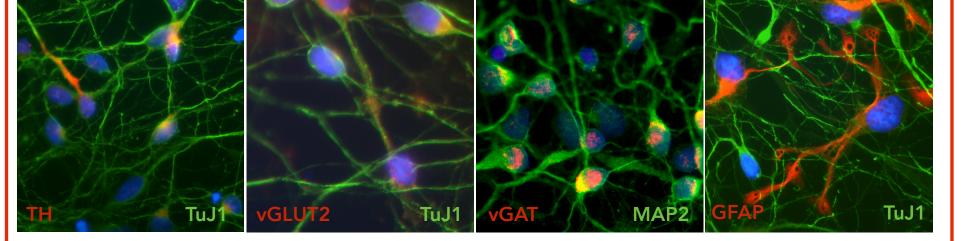
Calcium Oscillations in CNS.4U hiPSC-derived neurons detectable by Hamamatsu µCELL using Cal520 Calcium Dye



METHODS

<u>Calcium transcient flux</u>

- Instrument- Hamamatsu FDSS/µCell, 384 format
- Density- 30,000 cells/well
- Buffer Ncardia Ca Oscillations buffer



Ncardia neuron composition ideal for high throughput screening, toxicity, & seizure assessment studies. CNS.4U hiPSC-derived neurons from Ncardia represent a broad population of CNS neurons & astrocytes in a ready-to-use coculture format. Staining performed for functional markers is a standard quality control metric at Ncardia. Confocal microscopy was performed by Colin Eddington at MIT. CNS.4U show complex synaptic formation and astrocytic support.

MEA

- Instrument Axion Maestro System
- Density 3.6-7.2 x 10^4 cells were seeded per well in 3 µl droplets
- Buffer- Neuro.4U Basal Medium

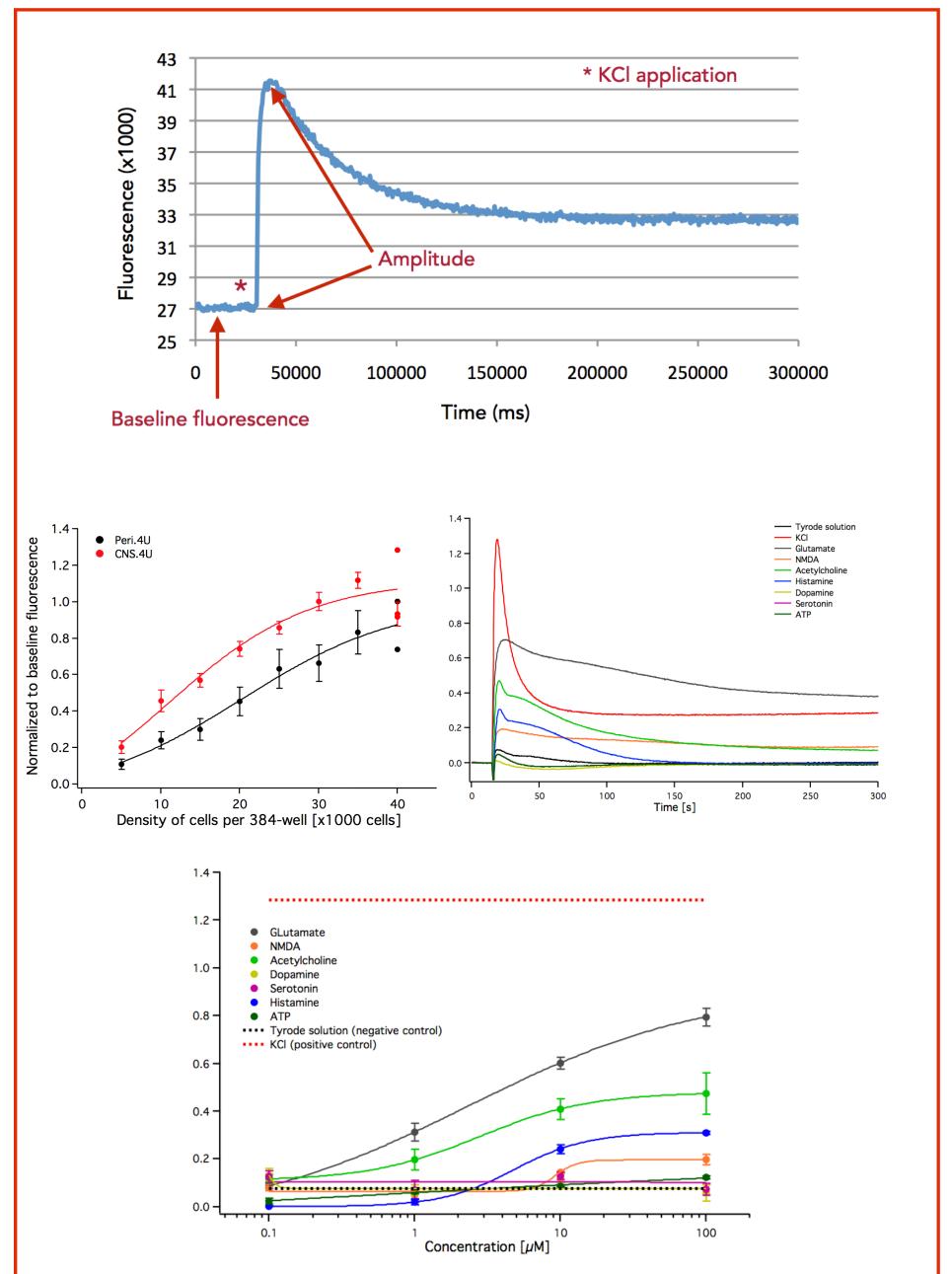
Baseline

Picrotoxin

- MEA recordings were performed with cells that had been cultured for up to 3 weeks (Seizure1.4U) or up to 8 weeks (Seizure2.4U and Seizure3.4U)

Calcium Oscillations in CNS.4U labeled with Cal520 Calcium Dye Cal520 is the calcium dye of choice by Ncardia when detecting neurotransmitter induced calcium oscillations in CNS.4U (3x10^4 cells/well in 384 well plate at 33 days in vitro). *Comparison calcium dye image from AATBio.com using CHO cells.

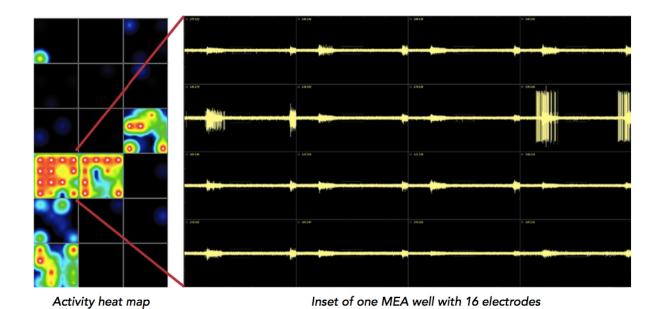
Next Frontier: HTS neurotransmitter screening of calcium oscillations in CNS.4U hiPSC-derived neurons



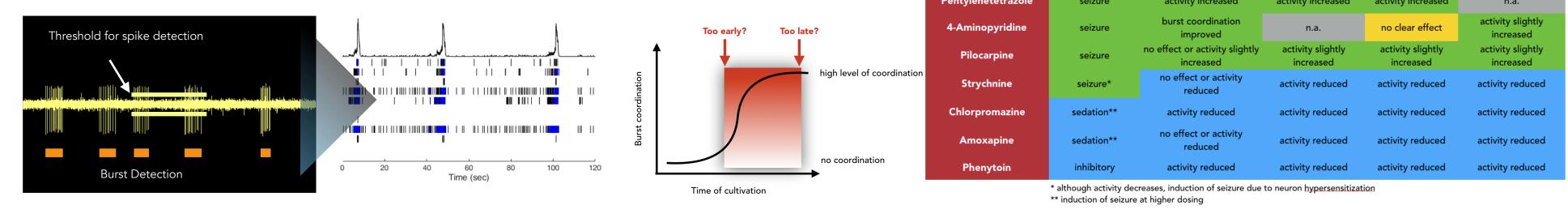
- Ca Dye: Cal520 (AAT Bio)
- Recording time: Day 33 in vitro post-thaw
- Drugs were diluted in medium and applied as single dose or cumulatively in increasing concentrations. During drug application, 10% of the bath solution was replaced with a 10-fold concentrated drug solution

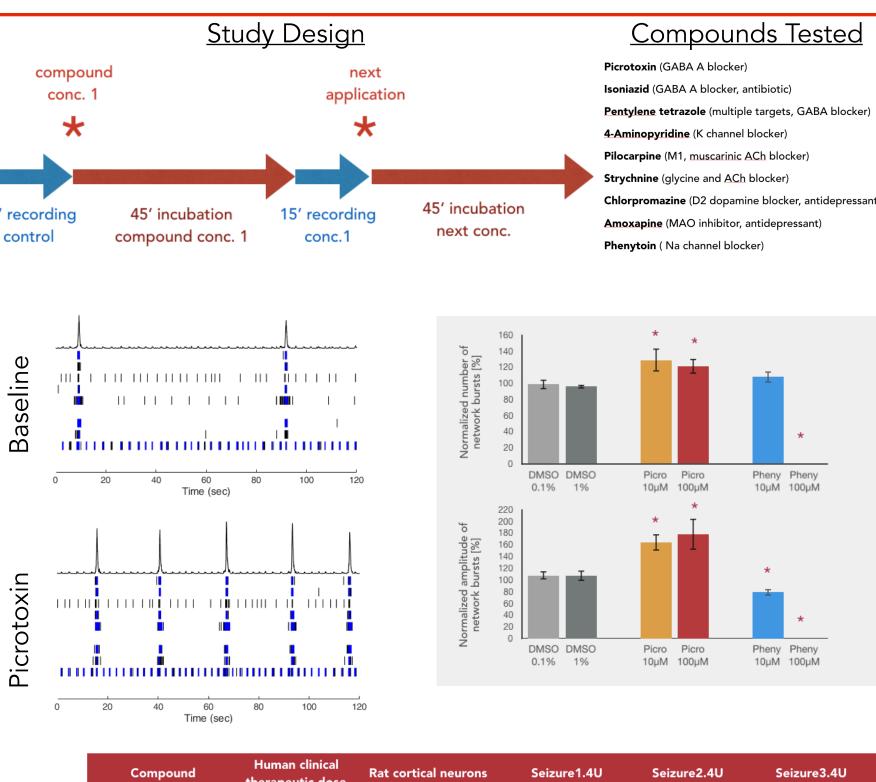
Pro-seizurogenic Drug Liability Assessment using 3 Different hiPSC-derived neural co-cultures for the HESI Neutox Consortium

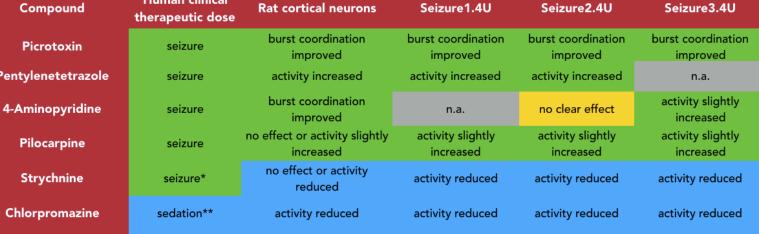




	Peri.4U	Development Model	CNS.4U
cell types	50 % Glutamate + 50% GABA	Largely TH+ (60% after 2 weeks)	30% Glu + 30% GABA + 10 % TH+ + 10% Astro
differentiated	protocol 1	protocol 2	protocol 3
age at freezing	24 days	18 days	24 days
cultivation time after thawing	7-14	> 18	> 24
Astro.4U added	15%	15%	0% (~10% inherent)
respond to	Glutamate, NMDA,		Glutamate, NMDA,
(MPC, Ca-transients)	GABA, ACh		GABA, ACh, Histamine







Early observations of neurotransmitter-induced calcium oscillations in CNS.4U Warning! Optimization is still required! - e.g. addition of glycine to improve NMDA currents. pH changes due to calcium dye and incubator are also complicating factors. A GCaMP6f calcium sensor (expressed via Xpress.4U) will also be explored to allow for long-term experiments and to help reduce calcium dye toxicity.

Ncardia hiPSC-derived neuron population Seizure 3.4U on Axion Maestro system detects pro-seizurogenic activity of Picrotoxin and reduced seizurogenic activity of Phenytoin Microelectrode array (MEA) technology offered by Axion in a 16 electrode/well format in combination with Ncardia hiPSC-derived neuron protocol Seizure 3.4U offers a translatable replacement for detecting pro-seizurogenic activity of known pharmacological agents in comparison to Human clinical data and rat cortical neuron models.

COOPERATION PARTNERS

PHOTON IS OUR BUSINESS

CONCLUSIONS

- Large lot numbers and stringent (and MEA based functional) QC parameters by Ncardia results in high quality hiPSC-derived neuronal cells ideal for translatable large scale studies and guarantees robust electrical activity
- Experiments reveal suitability of Ncardia hiPSC-derived neurons (+ astrocytes) for seizure liability assays given their long-term synchronous network activity and reactivity to seizure-active or -suppressive compounds
- The high acquisition rate camera on the Hamamatsu FDSS/µCell detects calcium flux in neurotransmitter stimulated hiPSCderived neurons labeled with Cal520 thus providing a new translatable neuron screening system for high throughput studies in up to 384 well format for pharmacology safety studies. This may present a novel screening strategy for seizure activity as well.



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